Multiple genetic analyses to investigate the polymorphisms of Chinese Mongolian population with an efficient short tandem repeat panel

Aim To determine allele frequencies and forensic statistics of 22 autosomal short tandem repeat loci in Chinese Mongolian population.

Methods Blood specimens were collected from 134 unrelated healthy Mongolian individuals, and 22 short tandem repeat loci were co-amplified and genotyped. Allele frequencies and forensic parameters were calculated, and population genetic differences were analyzed among Mongolian population and other eight Chinese populations: Northern Han, Guangdong Han, Chengdu Han, Xinjiang Hui, Xinjiang Uygur, Hainan Li, Qinghai Tibetan, and Hainan Han.

Conclusion The combined application of these 22 loci could be useful for forensic purposes in the Mongolian population. Mongolian population had smaller genetic distances from the populations in northern China (Northern Han, Xinjiang Uygur, and Xinjiang Hui) than from the populations in Hainan province (Hainan Han and Hainan Li populations).

Yating Fang¹, Tong Xie¹, Qiong Lan¹, Xiaoye Jin^{2,3}, Yuxin Guo^{2,3}, Yongsong Zhou¹, Jiangwei Yan⁴, Bofeng Zhu^{1,2,3}

¹School of Forensic Medicine, Southern Medical University, Guangzhou, China

²Key Laboratory of Shaanxi Province for Craniofacial Precision Medicine Research, College of Stomatology, Xi'an Jiaotong University, Xi'an, China

³College of Medicine & Forensics, Xi'an Jiaotong University Health Science Center, Xi'an, China

⁴CAS Key Laboratory of Genome Sciences and Information, Beijing Institute of Genomics, Chinese Academy of Sciences, Beijing, China

Received: June 13, 2018 Accepted: May 13, 2019

//ceepred.///d) //0/201

Correspondence to: Bofeng Zhu Department of Forensic Genetics, School of Forensic Medicine Southern Medical University Guangzhou, 510515 Guangdong, PR China zhubofeng7372@126.com

Xinjiang province is an ethnic autonomous region in the northwest of China, bordering Russia, Pakistan, India, Kazakhstan, Tajikistan, Kyrgyzstan, Afghanistan, and Mongolia. Historically, the province was home to an important route on the Silk Road, functioning as China's gateway to the west. It is inhabited by 47 ethnic groups, including Mongolians, which mainly inhabit the Bayingol Mongol Autonomous Prefecture, Hoboksar Mongol Autonomous County, and Bortala Mongol Autonomous Prefecture. Besides these Chinese regions, Mongolians also inhabit Mongolia and parts of Russia. They are predominantly shamanist and speak a language from the Mongolian group of Altaic family.

A short tandem repeat (STR) is a train of repetitive base sequences on the DNA strand. STR genetic polymorphisms can be analyzed by measuring the exact number of repeating units on the DNA. The novel panel used in this study is a STR genotyping system based on capillary electrophoresis analysis with 5-color fluorescence labeling, which encompasses Amelogenin gene and 22 autosomal STR loci: D1S1656, D2S1338, D3S3045, D4S2366, D5S2500, D6S477, D7S3048, D8S1132, D9S925, D10S1435, D11S2368, D12S391, D13S325, D14S608, D15S659, D16S539, D17S1290, D18S535, D19S253, D20S470, D21S1270, and D22-GATA198B05. The 22 loci are distributed in 22 pairs of autosomes, and their amplified fragments are less than 450 bp.

This panel was validated by a previous study (1), which assessed its sensitivity, accuracy, precision, stability, stutter percentage, peak height ratio, and species specificity. It was used to analyze allelic distribution in Northern Han (2), Southern Han (3), Chengdu Han (4), Hainan Li (5), Xinjiang Hui (6), and Xinjiang Uygur (7). In addition, detailed sequence information of these 22 loci was studied by Phillips et al (8). However, it is unknown whether these 22 STR loci are suitable for forensic application in Xinjiang Mongolian population. Based on the published findings, we hypothesized that the 22 loci had high genetic polymorphisms in Xinjiang Mongolian population and could be applied in this population for individual identification and paternity testing. To test these hypotheses, our study determined the allele frequencies of 22 STRs in Chinese Xinjiang Mongolian population, evaluated the system effectiveness of these 22 loci for individual identification and paternity testing in this population, and compared the findings with other reference populations.

MATERIALS AND METHODS

Material

This observational population genetics study was conducted in June 2017 at the Xi'an Jiaotong University. A total of 134 peripheral blood samples of volunteer unrelated healthy Mongolian individuals (90 women and 44 men) were collected from Chinese Xinjiang Uygur Autonomous Region and saved in the form of a paper blood collection card. Individuals were considered eligible if they met the following criteria: (i) there were no blood relationships between them, (ii) they all lived in Xinjiang Uygur Autonomous Region for over three generations, (iii) and there was no migration in their family history. Informed consent for study participation and data presentation was obtained from all volunteers before sampling. The study was approved by the Ethics Committee of the Institute for Xi'an Jiaotong University (Approval No. XJTULAC201, Nov 7, 2013).

DNA analysis

After the extraction of genomic DNA using the Chelex-100 method (9), Amelogenin gene locus and 22 STR loci were co-amplified using the Microreader 23sp ID kit (Suzhou Microread Genetics, Suzhou, China) on the GeneAmp PCR 9700 thermocycler (Applied Biosystems, Foster City, CA, USA) with 25 µL reaction volume. The amplified products were isolated and detected by capillary electrophoresis using the ABI PRISM 3130XL Genetic Analyzer (Applied Biosystems) with reference to internal lane standard Org500 (including 14 different length fragments: 50, 75, 100, 139, 150, 160, 200, 300, 340, 350, 400, 450, 490, 500 bp). Capillary electrophoresis results were analyzed using GeneMapper ID-X 1.3 software (Applied Biosystems). The 9947A was used as a positive control and DNA-free deionized water as a negative control. Our experiments strictly followed the internal control standards of the laboratory of Southern Medical University (Guangzhou, Guangdong, China).

Statistical analysis

The Hardy-Weinberg equilibrium (HWE) (10) of the 22 autosomal STR loci was tested by using Modified Powerstats software v. 1.2 (11), which was also used to compute allele frequencies and forensic parameters of each locus, ie, power of discrimination (PD), matching probability (MP), power of exclusion (PE), observed heterozygosity (Ho), and polymorphic information content (PIC). The expected heterozygosity (He) for each locus was calculated by Arle-

TABLE 1. The allele frequencies and forensic parameters for 22 autosomal short tandem repeat loci in Xinjiang Mongolian population (n = 134)*	Alleles D151656 D251338 D353045 D452366 D552500 D65477 D753048 D851132 D95925 D1051435 D1152368 D125391 D135325 D145608 D155659 D165539 D1751290 D185535 D195253 D2054
---	--

Alleles	Alleles D1S1656 D2S1338 D3S3045 D4S2366 D5S2500 D6S477 D7S3048 D8S1132	2S1338 D3	S3045 D.	4S2366 D)5S2500 l	D6S477 [07S3048			D10S14351	D1152368	D12S391 l	D9S925 D10S1435 D11S2368 D12S391 D13S325 D14S608 D15S659 D16S539 D17S1290 D18S535 D19S253 D20S470 D21S1270 D22-GATA198B05	3 D15S659	D16S539	D1751290	D1855351	D19S253 [020S470 E	D2151270D22	2-GATA198B05
9													0.0410						0.0224		
7										0.0075			0.1754					0.1679			
00				0.0037						0.0187			0.0149		0.0149			0.0112	0.0037		
6		0	0.2761	0.2948	0.0037								0.1194		0.3022		0.1940	0.0075	0.0187		
9.2						0.0037															
10	0.0037	0	0.0485	0.0746	0.0373	0.0149				0.0336			0.1978	0.0187	0.1045	0.0224	0.0373	0.0112	0.1231	0.3022	
10.2						0.0112															
11	0.0448	0	0.0485	0.2537	0.2910	0.0075			0.0112	0.1381			0.2201	0.1269	0.1903	0.0448	0.0224	0.0858	0.0373	0.1007	
11.2						0.0037															
12	0.0560	0	0.1119	0.1903	0.1716	0.0448			0.0075	0.3657			0.1828	0.2239	0.1903	0.0037	0.1157	0.3657	0.0672	0.0597	
12.2						0.0112															
12.3																			0.0037	0.0560	
13	0.1007	C	0.1978	0.0709	0.0485	0.2463				0.2313			0.0485	0.1157	0.1716		0.2687	0.2463	0.0970	0.1231	
13.3																			0.0037	0.0224	
14	0.0485	0	0.2239	0.0933	0.0485	0.1679			0.1082	0.1754				0.0597	0.0261	0.0149	0.2687	0.0858	0.1679	0.2351	0.0037
14.3																				0.0224	
15	0.2948	0.0037 0	0.0858	0.0187	0.2799	0.2276		0.0037	0.2761	0.0299	0.0037	0.0149		0.1940		0.2052	0.0821	0.0187	0.1940	0.0709	
15.3	0.0112																				
16	0.2612	0.0075 0	0.0075		0.1007	0.2052		0.0075	0.3022		0.0149		0.0149	0.1642		0.3582	0.0112		0.1381	0.0075	0.0597
16.3	0.0224								0.0037												
17	0.0672	0.0821			0.0187	0.0373	0.0075	0.0821	0.1866		0.1791	0.1231	0.0373	0.0746		0.1493			0.0896		0.1157
17.3	0.0560								0.0037			0.0112									
18		0.1045				0.0075	0.1306	0.2090	0.0933		0.1157	0.2500	0.0448	0.0224		0.1418			0.0149		0.1007
18.3	0.0187																				
19	0.0037	0.1791				0.0075	0.0858	0.1530	0.0075		0.1455	0.1530	0.2425			0.0522			0.0149		0.1045
19.3	0.0075																				
20		0.1045				0.0037	0.1231	0.0933			0.1381	0.1828	0.2724			0.0037			0.0037		0.1007
20.3	0.0037																				
21		0.0187					0.1082	0.1343			0.2463	0.0933	0.2052								0.2761
22		0.0522					0.0970	0.1269			0.1045	0.1007	0.1194			0.0037					0.1642
23		0.1455					0.1418	0.1530			0.0373	0.0597	0.0373								0.0634
24		0.1604					0.1791	0.0336			0.0112	0.0112	0.0261								0.0112
25		0.1045					0.1157	0.0037			0.0037										
26		0.0224					0.0037														
27		0.0075					0.0075														
28		0.0075																			
MP	0.0564	0.0330 0	0.0635	0.0792	0.0740	0.0661	0.0342	0.0441	0.0828	0.0972	0.0503	0.0478	0.0716 0.0626	0.0481	0.0783	0.0933	0.0764	0.1016	0.0363	0.0581	0.0468
PD	0.9436	0.9670 0	0.9365	0.9208	0.9260	0.9339	0.9658	0.9559	0.9172	0.9028	0.9497	0.9522	0.9284 0.9374	0.9519	0.9217	0.9067	0.9236	0.8984	0.9637	0.9419	0.9532
PIC	0.7980	0.8654 0	0.7838	0.7631	0.7615	0.7872	0.8600	0.8425	0.7427	0.7252	0.8215	0.8238	0.7778 0.8069	0.8272	0.7648	0.7532	0.7663	0.7279	0.8627	0.7930	0.8274
PE	0.6527	0.8168 0	0.6817	0.5963	0.5555	0.6384	0.7260	0.8015	0.5422	0.5034	0.7111	0.6963	0.5291 0.7409	0.5963	0.5689	0.5555	0.6242	0.5689	0.8321	0.6102	0.6963
Ю	0.8284			0.7985	0.7761	0.8209		0.9030		0.7463	0.8582	0.8507	0.7612 0.8731	0.7985	0.7836	0.7761	0.8134	0.7836	0.9179	0.8060	0.8507
	10000				00001	0,100		0000				0		0							11000

Чe

quin software v. 3.5 (12), which was applied to determine whether there was a linkage disequilibrium (LD) (13) between these loci. The combined power of discrimination (CPD) and combined probability of exclusion (CPE) were calculated using the respective formulas:

$$\begin{split} \mathsf{CPD} &= 1 - (1 - \mathsf{DP}_1)(1 - \mathsf{DP}_2)(1 - \mathsf{DP}_3) \cdots (1 - \mathsf{DP}_k); \ \mathsf{CPE} = 1 - (1 - \mathsf{PE}_1) \\ (1 - \mathsf{PE}_2) &(1 - \mathsf{PE}_3) \cdots (1 - \mathsf{PE}_k). \end{split}$$

In these two formulas, k indicates the number of loci. The allele frequencies of 22 loci were compared between the

Mongolian and other reference populations by Arlequin software v. 3.5 (12). The STRUCTURE analysis was performed by using the STRUCTURE software v. 2.3.4 (14). Genetic distances (D_A) and fixation index (Fst) values in Mongolian and other populations were calculated with DISPAN (15) and Genepop software v. 4.0 (16), respectively. Heat maps were drawn by R software v. 3.4.3 (17) based on the D_A and Fst values, and phylogeny trees were drawn by MEGA software v. 6.0 (18) and Phylip software v. 3.69 (19-21). Principal components analysis (PCA) was performed by using and MVSP software v. 3.1 (22). All the used software is freely available.



FIGURE 1. Stacked histogram showing forensic parameters of 22 autosomal short tandem repeat loci in Xinjiang Mongolian population (n = 134).



FIGURE 2. Heat map (**A**) showing the pairwise genetic distances between Xinjiang Mongolian and eight other populations based on allele frequencies of 16 overlapping loci. Heat map (**B**) showing pairwise fixation index (Fst) values of Xinjiang Mongolian and six other populations based on the data of 16 overlapping loci.

RESULTS

Hardy-Weinberg equilibrium and linkage disequilibrium tests

The *P* values of 22 loci were all greater than 0.05, which meant that none of these loci deviated from the HWE (Table 1). When the loci were tested for linkage disequilibrium, the *P* values of 22 out of 231 pairwise loci were less than 0.05. After applying the Bonferroni correction (23), the adjusted significance level was 0.0002 (0.05/231), which indicated that there was no linkage disequilibrium between these loci (Supplementary Table 1). In other words, these loci were independent from each other.

Allele frequencies and forensic statistical parameters

A total of 227 alleles were detected at these 22 loci (Table 1). The highest number of alleles was detected at the D20S470 locus (16 alleles) and the lowest number at the D16S539 locus (7 alleles). The highest allele frequency was 0.3657 at two loci and the lowest was 0.0037 at 12 loci.

 He values from 0.7634 (D10S1435) to 0.8810 (D2S1338). The heterozygosity of all the loci was greater than 0.7. These results indicate that these loci have high discrimination power in Xinjiang Mongolian population (24,25). The minimum value of PIC was 0.7252 (D10S1435), indicating that all these loci were highly polymorphic. The PE values ranged from 0.5034 (D10S1435) to 0.8321 (D20S470). The CPE value was 0.999999999566925, which met the Forensic Science DNA Parentage Test Specification issued by the Ministry of Public Security of the People's Republic of China in 2011 (GAT965-2011) that the CPE of the STR panel used in the triad paternity test should not be less than 0.9999 (Table 1) (Figure 1).

Population genetic analysis

We assessed the differences between Xinjiang Mongolian population and six reference populations in the allele frequencies of these 22 loci by using the analysis of molecular variance (Table 2). There were significant differences at 6, 8, 6, 8, 9, and 17 loci between Mongolian population and Northern Han (Hebei, Henan, Shaanxi) (2), Guangdong Han (3), Chengdu Han (4), Xinjiang Hui (6), Xinjiang Uygur (7), and Hainan Li (5) populations, respectively. In addition, we chose 16 loci as overlapping loci (except for D2S1338, D9S925, D12S391, D16S539, D20S470, and D21S1270) to compare the studied population with the Qinghai Tibetan (26) and Hainan Han (27) populations. Significant dif-



FIGURE 3. Phylogenetic tree (**A**) showing the relationships between Xinjiang Mongolian and eight other populations based on the results of genetic distance analysis. Phylogenetic tree (**B**) showing the relationships between Xinjiang Mongolian and eight other populations based on the allele frequencies of 16 overlapping loci.

Loci	Northern Han	Guangdong Han	Chengdu Han	Xinjiang Hui	Xinjiang Uygur	Hainan Li	Hainan Han	Qinghai Tibetan
D1S1656	0.6686	0.0968	0.0772	0.8338	0.0029	<0.001	< 0.001	0.4330
D2S1338	0.0186	0.0264	0.5024	0.0323	0.4565	0.1525		—
D3S3045	0.1173	0.0362	0.0723	0.2092	0.0479	< 0.001	0.0010	0.8397
D4S2366	0.0039	0.1232	0.0127	0.0029	0.1652	< 0.001	0.0029	0.0010
D5S2500	0.5171	0.3118	0.8993	0.5699	0.0968	0.4780	0.1476	0.6256
D6S477	0.0978	0.0968	0.1867	0.0254	0.0401	< 0.001	0.0244	0.1613
D7S3048	0.0821	0.3324	0.0313	0.1496	0.0557	<0.001	< 0.001	0.0039
D8S1132	< 0.001	0.0303	0.0166	0.0039	0.3099	<0.001	< 0.001	0.0010
D9S925	0.0538	0.0147	0.4291	0.2141	0.0694	0.1417	—	—
D10S1435	0.4702	0.4555	0.5816	0.4330	0.1271	0.5562	0.9394	0.5533
D11S2368	0.1916	0.0616	0.5748	0.0420	0.0372	0.0274	0.1369	0.1623
D12S391	0.2258	0.0127	0.1193	0.5142	0.0635	0.0098	_	—
D13S325	0.6061	0.5435	0.7380	0.6334	0.4311	0.0049	0.1281	0.4096
D14S608	0.0284	0.0156	0.0068	0.0205	< 0.001	0.0020	< 0.001	0.1144
D15S659	0.7185	0.0782	0.1183	0.6501	0.2483	< 0.001	0.0156	0.2434
D16S539	0.0362	0.1017	0.0117	0.0098	0.0010	< 0.001	_	—
D17S1290	0.1271	0.0362	0.0313	0.3558	0.2708	< 0.001	0.0010	0.1408
D18S535	0.7322	0.0411	0.8974	0.7175	0.0899	< 0.001	< 0.001	0.2718
D19S253	0.0215	0.1720	0.3157	0.1584	0.3529	0.0606	0.0020	0.6843
D20S470	0.0938	0.0547	0.3783	0.0156	0.0284	0.0039	—	_
D21S1270	0.1105	0.0880	0.3969	0.2297	0.0020	0.0108	—	_
D22-GATA198B05	0.1173	0.2669	0.4282	0.2835	0.0274	<0.001	0.0469	0.1593

TABLE 2. The P values of the locus-by-locus comparisons based on the allele frequencies in Xinjiang Mongolian and other populations

TABLE 3. The pairwise genetic distance values based on the allele frequencies of 16 loci in Xinjiang Mongolian and eight reference populations

Populations	Xinjiang Mongolian	Xinjiang Hui	Northern Han	Xinjiang Uygur	Qinghai Tibetan	Chengdu Han	Guangdong Han	Hainan Han
Xinjiang Hui	0.0113			- / 5				
Northern Han	0.0115	0.0035						
Xinjiang Uygur	0.0141	0.0105	0.0134					
Qinghai Tibetan	0.0163	0.0067	0.0073	0.0160				
Chengdu Han	0.0175	0.0077	0.0067	0.0180	0.0107			
Guangdong Han	0.0212	0.0116	0.0105	0.0208	0.0155	0.0121		
Hainan Han	0.0217	0.0117	0.0096	0.0206	0.0160	0.0099	0.0108	
Hainan Li	0.0379	0.0253	0.0242	0.0346	0.0305	0.0205	0.0214	0.0135

TABLE 4. The pairwise fixation index values based on allele frequencies of 16 loci in Xinjiang Mongolian and six reference populations

Populations	Xinjiang Mongolian	Xinjiang Hui	Northern Han	Xinjiang Uygur	Guangdong Han	Hainan Han
Xinjiang Hui	0.0014					
Northern Han	0.0015	0.0002				
Xinjiang Uygur	0.0023	0.0031	0.004			
Guangdong Han	0.0033	0.0025	0.0015	0.0046		
Hainan Han	0.0059	0.0046	0.0036	0.0072	0.0008	
Hainan Li	0.0138	0.012	0.0109	0.0133	0.0053	0.0033

ferences were observed between Mongolian and Qinghai Tibetan population at 12 loci, and between Mongolian and Hainan Han population at 3 loci.

The structure analysis of seven populations (Xinjiang Mongolian, Northern Han, Guangdong Han, Chengdu Han, Xinjiang Hui, Xinjiang Uygur, and Hainan Li) offered no evidence that they had different component distribution. Next, we used a series of bioinformatics methods to analyze the genetic relationships between the populations. D. between the populations were calculated based on allele frequencies of 16 overlapping loci. Fst values (a measure of genetic differentiation) between any two of seven populations (except for Chengdu Han and Qinghai Tibetan) were obtained to quantify the genetic relationships between different groups. $\mathsf{D}_{\scriptscriptstyle A}$ values ranged from 0.0035 to 0.0379 and Fst values from 0.0002 to 0.0138 (Table 3 and Table 4). Xinjiang Mongolian population had the smallest genetic distances from Xinjiang Hui (D, =0.0113, Fst=0.0014), Northern Han (D_{a} =0.0115, Fst=0.0015), and Xinjiang Uygur ($D_{A} = 0.0141$, Fst = 0.0023), and the greatest genetic distance from Hainan Li (D₄=0.0379, Fst=0.0138). Overall, the relationships among these populations were relatively close (Figure 2).

Based on D₄ values and allele frequencies we constructed two phylogenetic trees (Figure 3). The populations were divided into two sub-branches. Hainan Han and Hainan Li formed the first sub-branch and other populations formed the second one. In the second sub-branch, Xinjiang Mongolian and Xinjiang Uygur clustered together, followed by Qinghai Tibetan and Xinjiang Hui, and then clustered with the Han populations from different regions. PCA analysis also showed the aggregation of populations. Xinjiang Mongolian and Xinjiang Uygur gathered in the upper right corner, while Xinjiang Hui, Qinghai Tibetan, Northern Han, and Chengdu Han gathered in the lower right corner. In the lower left corner there were Guangdong Han and Hainan Han, while Hainan Li was far away from other populations (Figure 4). The results indicate that the genetic distances between Xinjiang Mongolian and populations in the northern regions of China (Northern Han, Xinjiang Uygur, and Xinjiang Hui) were even smaller. On the other hand, the distances from Hainan Li and Hainan Han popu-



FIGURE 4. Principal component analysis based on the allele frequencies of 16 short tandem repeat loci of Xinjiang Mongolian and eight other populations.

lations were large. This is consistent with the previous results (28).

DISCUSSION

Xinjiang Mongolians had high polymorphism at these 22 STR loci, which confirmed that the combined application of these loci was appropriate for individual identification and paternity testing in this population. In conclusion, our hypotheses were confirmed. Population genetic analysis revealed the genetic relationships between Xinjiang Mongolian and other eight Chinese populations.

With the advancement of science and technology, several new technologies and genetic markers, like next-generation sequencing and single nucleotide polymorphisms, have become widely used. However, due to the lack of databases for new genetic markers, STR typing is still used in the forensic practice. The Federal Bureau of Investigation laboratory in 1997 selected 13 autosomal STRs as core loci of Combined DNA Index System (CODIS) (29), which was in 2017 expanded to 20 STRs (30). Commercially most available STR kits are based on these core loci (31-33). In recent years, these core loci have been complemented by more and more new non-CODIS loci to gain additional genetic information and further improve the discriminatory power (8). Among the studied 22 loci, only four were CODIS loci (D1S1656, D2S1338, D12S391, and D16S539) (30), which increased DNA marker coverage in forensic application. Among other 18 loci, as far as we know, D9S925, D20S470, and D21S1270 are new loci adopted only by this system, which are not included in other commercial kits (8). In fact, newly-adopted STRs should be cautiously used. To verify whether new STRs are suitable for forensic application, it is necessary to perform their detailed genomic characterization and conduct a number of population surveys (8). Detailed studies of the gene sequence information of these 22 loci, especially the newly adopted non-CODIS STRs, as well as the validation studies on the sensitivity, accuracy, and species specificity of this new panel have been performed (1,8). Polymorphisms of these loci in Han, Li, Hui, and Uygur populations in some regions of China have also been reported (2-7). On the basis of these studies, we analyzed the genetic polymorphism of these loci in Mongolian population in Xinjiang.

The sample size in this study (n = 134) was based on previous studies (34,35). Given that the HWE tests were the basis of the population genetics study, the *post hoc* power analysis of HWE tests was performed by R version 3.6.0 (36) (Supplementary Table 2). The results showed that 17 out of 22 loci had power greater than 0.8. Although *post hoc* power analysis has some limitations in sample size evaluation (37), it indicated that in future studies we may need to increase the sample size to obtain more genetic polymorphism information about the other five loci (especially D3S3045 and D9S925 with power less than 0.5).

Our study confirmed the forensic applicability of these 22 loci in Xinjiang Mongolian population. However, due to the small number of population data on this new system (currently only eight populations have available data), the genetic relationships have to be interpreted in light of certain limitations. In order to further conduct population research and explain the origin of Mongolians, the genetic characteristics of these 22 loci should be evaluated in other populations and genetic characteristics of Mongolians at other loci should be assessed.

Funding The study was supported by the National Natural Science Foundation of China (NSFC, project number 81525015, 81373248).

Ethical approval given by the Ethics Committee of the Institute for Xi'an Jiaotong University (Approval No. XJTULAC201, Nov 7, 2013).

Declaration of authorship BFZ conceived and designed the study; YTF, TX, QL, XYJ, and YXG acquired the data; QL, XYJ, YKG, YSZ, and JWY analyzed and interpreted the data; YTF, TX, QL, XYJ, and YXG drafted the manuscript; YTF, YSZ, JWY, and BFZ critically revised the manuscript for important intellectual content; all authors gave approval of the version to be submitted; all authors agree to be accountable for all aspects of the work.

Competing interests All authors have completed the Unified Competing Interest form at www.icmje.org/coi_disclosure.pdf (available on request from the corresponding author) and declare: no support from any organization for the submitted work; no financial relationships with any organizations that might have an interest in the submitted work in the previous 3 years; no other relationships or activities that could appear to have influenced the submitted work.

References

- Li J, Luo H, Song F, Zhang L, Deng C, Yu Z, et al. Validation of the Microreader[™] 23sp ID system: A new STR 23-plex system for forensic application. Forensic Sci Int Genet. 2017;27:67-73. Medline:27992828 doi:10.1016/j.fsigen.2016.12.005
- 2 Xie B, Chen L, Yang Y, Lv Y, Chen J, Shi Y, et al. Genetic distribution of 39 STR loci in 1027 unrelated Han individuals from Northern China. Forensic Sci Int Genet. 2015;19:205-6. Medline:26264960 doi:10.1016/j.fsigen.2015.07.019
- 3 Liu QL, Chen ZX, Chen CG, Lu DJ. Genetic polymorphism of 22 autosomal STR markers in a Han population of Southern China. Forensic Sci Int Genet. 2016;24:e14-6. Medline:27421761 doi:10.1016/j.fsigen.2016.06.017
- 4 Li JN, Luo HB, Song F, Hou YP. Genetic polymorphisms of 22 autosomal STR loci in Chinese Han population. Forensic Sci Int Genet. 2015;5:e45-7. doi:10.1016/j.fsigss.2015.09.018
- 5 Chen J, Xie B, Yang Y, Yang M, Liu C, Lv Y, et al. Genetic variability and forensic efficiency of 39 microsatellite loci in the Li ethnic

group from Hainan Island in the South China Sea. Ann Hum Biol. 2017;44:467-74. Medline:27670224 doi:10.1080/03014460.2016.12

6 Fang Y, Guo Y, Xie T, Jin X, Lan Q, Zhou Y, et al. Forensic molecular genetic diversity analysis of Chinese Hui ethnic group based on a novel STR panel. Int J Legal Med. 2018;132:1297-9. Medline:29582135 doi:10.1007/s00414-018-1829-1

41300

- 7 Song F, Li J, Zhang L, Luo H. Population data for 22 autosomal STR loci in the Uygur ethnic minority. Int J Legal Med. 2017;131:651-2. Medline:27604533 doi:10.1007/s00414-016-1451-z
- 8 Phillips C. A genomic audit of newly-adopted autosomal STRs for forensic identification. Forensic Sci Int Genet. 2017;29:193-204. Medline:28463807 doi:10.1016/j.fsigen.2017.04.011
- 9 Walsh PS, Metzger DA, Higuchi R. Chelex 100 as a medium for simple extraction of DNA for PCR-based typing from forensic material. Biotechniques. 1991;10:506-13. Medline:1867860
- 10 Emigh TH. A comparison of tests for Hardy-Weinberg equilibrium. Biometrics. 1980;36:627-42. Medline:25856832 doi:10.2307/2556115
- 11 Zhao F, Wu XY, Cai GQ, Xu CC. The application of Mdified-Powerstates software in forensic biostatistics [in Chinese]. Chin J Forensic Med. 2003;18:297-8.
- 12 Excoffier L, Lischer HE. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. Mol Ecol Resour. 2010;10:564-7. Medline:21565059 doi:10.1111/j.1755-0998.2010.02847.x
- 13 Slatkin M. Linkage disequilibrium–understanding the evolutionary past and mapping the medical future. Nat Rev Genet. 2008;9:477-85. Medline:18427557 doi:10.1038/nrq2361
- Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multilocus genotype data. Genetics. 2000;155:945-59. Medline:10835412
- 15 Tatsuya O. DISPAN Software. The Pennsylvania State University.
 1993 Available from: http://www.personal.psu.edu/nxm2/software.
 htm. Accessed: December 12, 2016.
- 16 Rousset F. genepop'007: a complete re-implementation of the genepop software for Windows and Linux. Mol Ecol Resour. 2008;8:103-6. Medline:21585727 doi:10.1111/j.1471-8286.2007.01931.x
- 17 Gentleman R, Ihaka R, Bates D, Chambers J, Dalgaard P, Hornik K, et al. R language and environment. Version 3.4.3 [software]. Available from: https://www.r-project.org/. Accessed: December 22, 2017.
- 18 Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol. 2011;28:2731-9. Medline:21546353 doi:10.1093/molbev/msr121
- Nei M, Tajima F, Tateno Y. Accuracy of estimated phylogenetic trees from molecular data. II. Gene frequency data. J Mol Evol. 1983;19:153-70. Medline:6571220 doi:10.1007/BF02300753

- 20 Takezaki N, Nei M. Genetic distances and reconstruction of phylogenetic trees from microsatellite DNA. Genetics. 1996;144:389-99. Medline:8878702
- Alanzi ARA, Degnan JH. Inferring rooted species trees from unrooted gene trees using approximate Bayesian computation. Mol Phylogenet Evol. 2017;116:13-24. Medline:28780022 doi:10.1016/j.ympev.2017.07.017
- 22 MVSP. A Multivariate Statistical Package for Windows. Version 3.1 [software]. Kovach Computing Services. Available from: https:// mvsp-a-multivariate-statistical-package.apponic.com/. Accessed: December 12, 2016.
- 23 Curtin F, Schulz P. Multiple correlations and Bonferroni's correction.
 Biol Psychiatry. 1998;44:775-7. Medline:9798082 doi:10.1016/ S0006-3223(98)00043-2
- Wang YL, Sheng X, Li M, Chen YL, Lin Y, Chen LQ. Forensic application of HuaxiaTM platinum kit [in Chinese]. Fa Yi Xue Za Zhi.
 2017;33:129-35. Medline:29231017
- 25 Gill P. DNA as evidence–the technology of identification. N Engl J Med. 2005;352:2669. Medline:15987914 doi:10.1056/ NEJMp048359
- Gou H, Zhou J, Zhang Y, Wang C, Ma X, Ma H, et al. Allele frequency distribution of 21 forensic autosomal STR loci of Goldeneye DNA ID 22NC Kit in Chinese Tibetan group. Forensic Sci Int Genet. 2016;22:e21-4. Medline:26961075 doi:10.1016/j.fsigen.2016.02.008
- Fu X, He J, Guo J, Liu Y, Wang Z, Lv J, et al. Genetic distribution of 21
 STR loci in the Chinese Han population from Hainan Island using
 Goldeneye DNA ID 22NC Kit. Forensic Sci Int Genet. 2017;29:e19 20. Medline:28462809 doi:10.1016/j.fsigen.2017.04.014
- 28 Tao R, Zhang J, Bian Y, Dong R, Liu X, Jin C, et al. Investigation of 12 X-STR loci in Mongolian and Eastern Han populations of China with comparison to other populations. Sci Rep. 2018;8:4287. Medline:29523825 doi:10.1038/s41598-018-22665-3
- Butler JM. Genetics and genomics of core short tandem repeat loci used in human identity testing. J Forensic Sci. 2006;51:253-65.
 Medline:16566758 doi:10.1111/j.1556-4029.2006.00046.x
- Hares DR. Selection and implementation of expanded CODIS core loci in the United States. Forensic Sci Int Genet. 2015;17:33-4.
 Medline:25797140 doi:10.1016/j.fsigen.2015.03.006
- 31 Builes JJ, Trejos D, Suárez D, Moreno S, Siza L, Acevedo M, et al. New alternative for human identification. Investigator IDplex Kit – QIAGEN® reproducibility: Latin American interlaboratory study. Forensic Sci Int Genet. 2011;3:e83-4. doi:10.1016/j. fsigss.2011.08.041
- 32 Mattayat D, Kitpipit T, Phetpeng S, Asawutmangkul W, Thanakiatkrai P. Comparative performance of AmpFLSTR[®] Identifiler[®] Plus PCR amplification kit and QIAGEN[®] Investigator[®] IDplex Plus kit. Sci Justice. 2016;56:468-74. Medline:27914554 doi:10.1016/j.scijus.2016.06.008
- 33 Tan JYY, Tan YP, Ng S, Tay AS, Phua YH, Tan WJ, et al. A preliminary evaluation study of new generation multiplex STR kits comprising

СМ

of the CODIS core loci and the European Standard Set loci. J Forensic Leg Med. 2017;52:16-23. Medline:28797659 doi:10.1016/j. jflm.2017.07.017

- Hale ML, Burg TM, Steeves TE. Sampling for microsatellite-based population genetic studies: 25 to 30 individuals per population is enough to accurately estimate allele frequencies. PLoS One. 2012;7:e45170. Medline:22984627 doi:10.1371/journal. pone.0045170
- Gao Y, Li SB. Effects of sample size on the observed number of allele of 9 STR loci with various genetic data [in Chinese]. Hereditas. 2008;30:313-20. Medline:18331999 doi:10.3724/ SPJ.1005.2008.00313
- R version 3.6.0: The R Project for Statistical Computing. Version
 3.6.0 [software]. Available from: https://www.r-project.org/
 Accessed: April 26, 2019.
- 37 Levine M, Ensom MH. Post hoc power analysis: an idea whose time has passed? Pharmacotherapy. 2001;21:405-9. Medline:11310512 doi:10.1592/phco.21.5.405.34503